

# Genome-wide discovery of modulators of transcriptional interactions in human B lymphocytes



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Ilya Nemenman

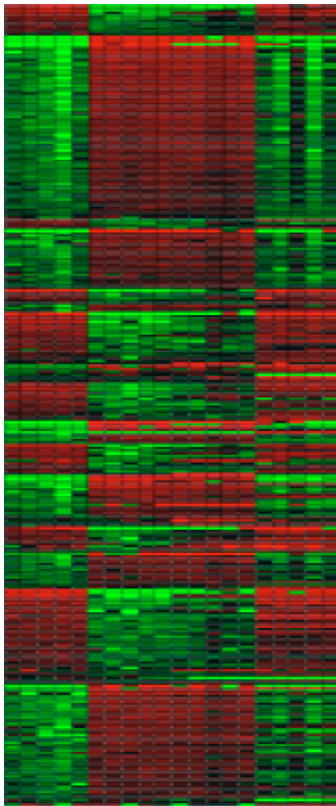
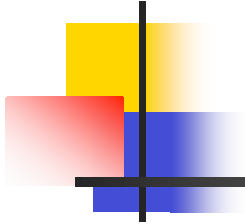
(JCSB/Columbia → CCS-3/LANL & SFI)

Kai Wang, Nilanjana Banerjee,

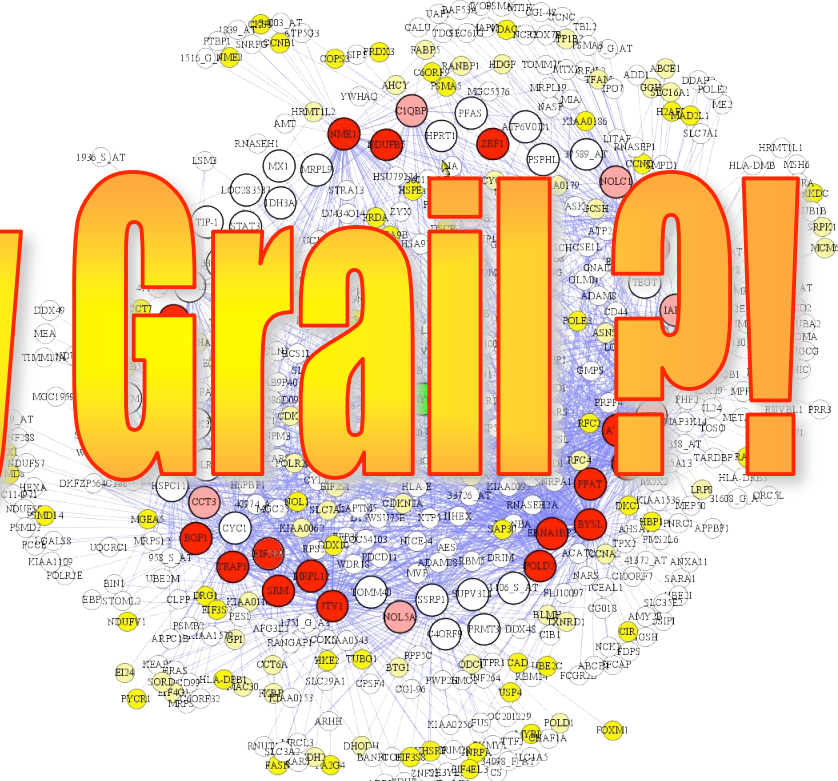
Adam Margolin, Andrea Califano

(JCSB/Columbia)

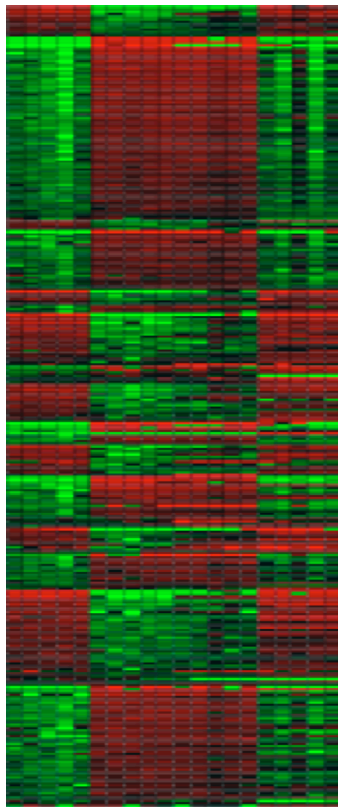
# Reconstructing cellular interactions



Holy Grail!?!?

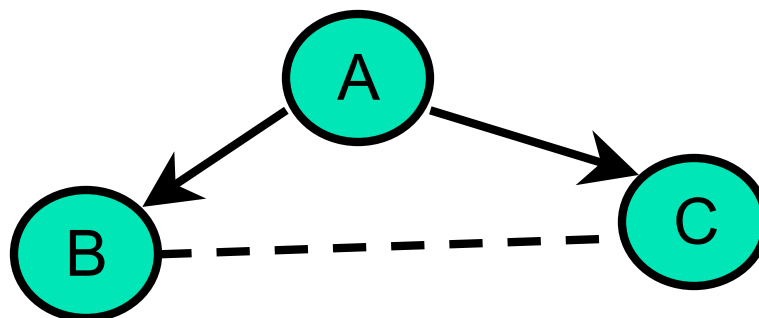


# Reconstruction algorithms: The curse of “percent correct”

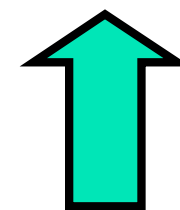


Small data requirements  
Robustness to fluct.  
Computational complexity  
Conditional interactions  
Reparam inv., non-param.

Irreducibility



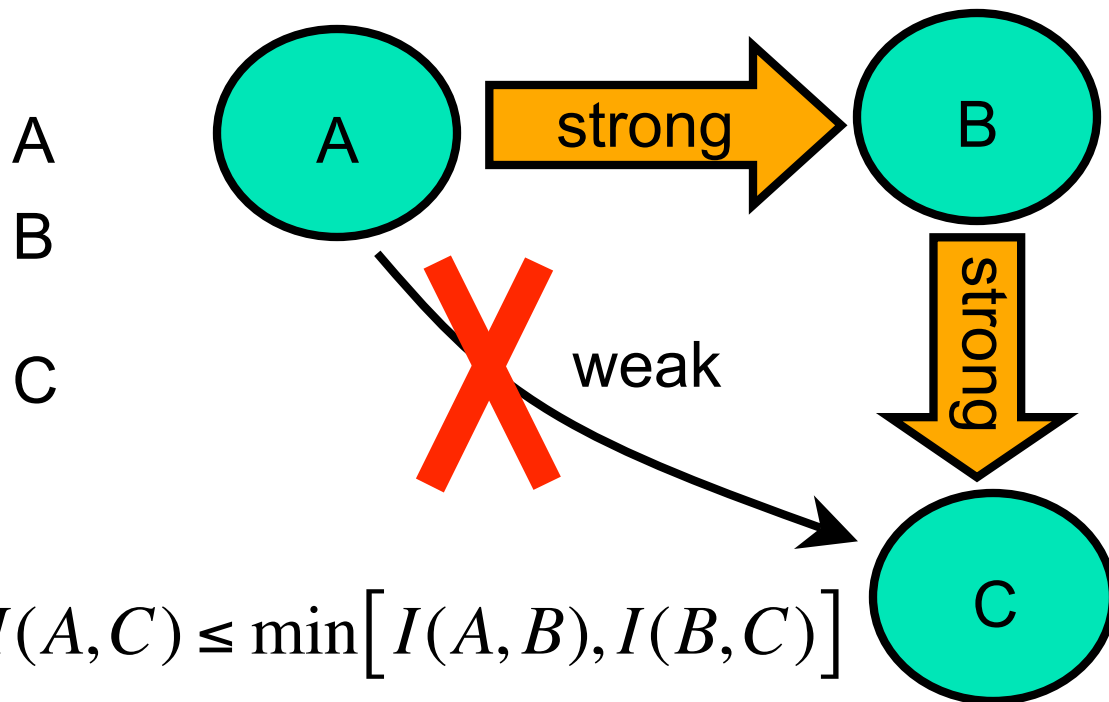
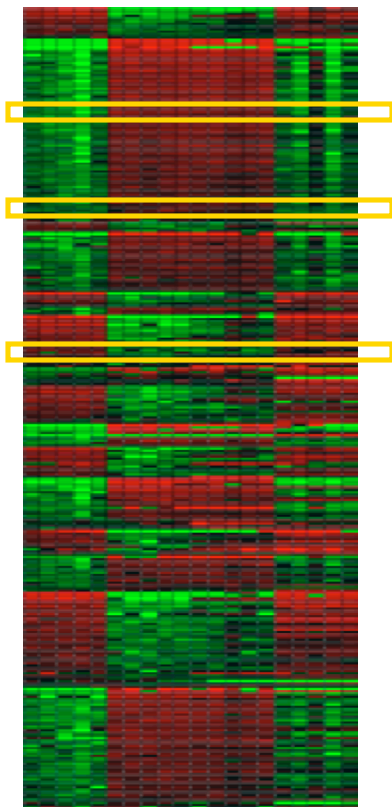
Stat	Co	GM	Biochem.
✗✓	✓	✗✓	✗
✓	✓	✗✓	✗
✗	✓	✗	✗✓
✓	✗✓	✓	✗✓
✗✓	✗✓	✗✓	✓
✓	✗	✓	✗



Influenciomics

# ARACNE

(Data Processing Inequality, DPI)



Reparm. inv.; small sample; low complexity.

**Performance?**





# Performance:

## No false positives

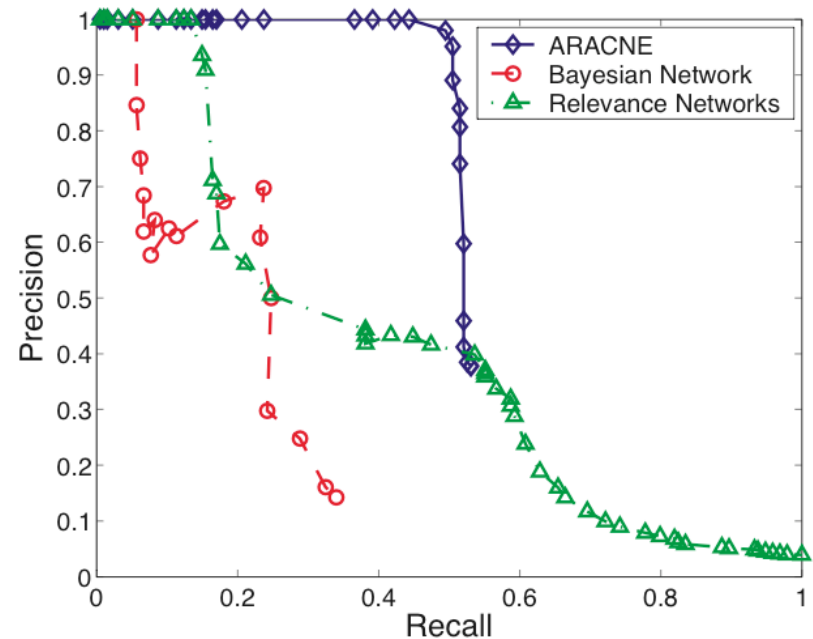
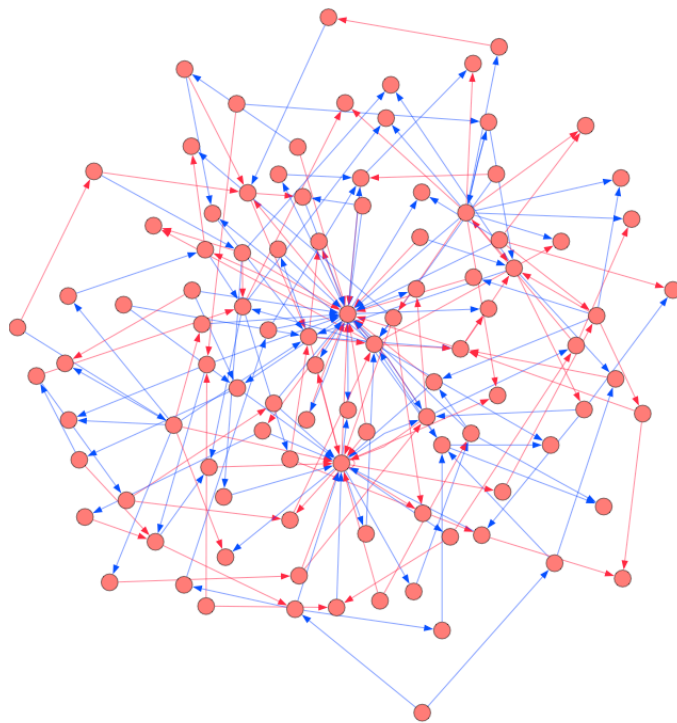
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Theorem 1. If MIs can be estimated with no errors, then ARACNE reconstructs the underlying interaction network exactly, provided this network is a tree and has only pairwise interactions.

Theorem 2. The Chow-Liu maximum mutual information tree is a subnetwork of the network reconstructed by ARACNE.

Theorem 3. Some sparseness/loopiness assumptions -- no false positives (no false negatives under stronger conditions).

# Synthetic networks



$$\frac{dx_i}{dt} = a_i \prod_j \frac{I_{0,j}^{v_j}}{I_j^{v_j} + I_{0,j}^{v_j}} \prod_j \left( 1 + \frac{A_{0,j}^{v_j}}{A_j^{v_j} + A_{0,j}^{v_j}} \right) - b_i x_i$$



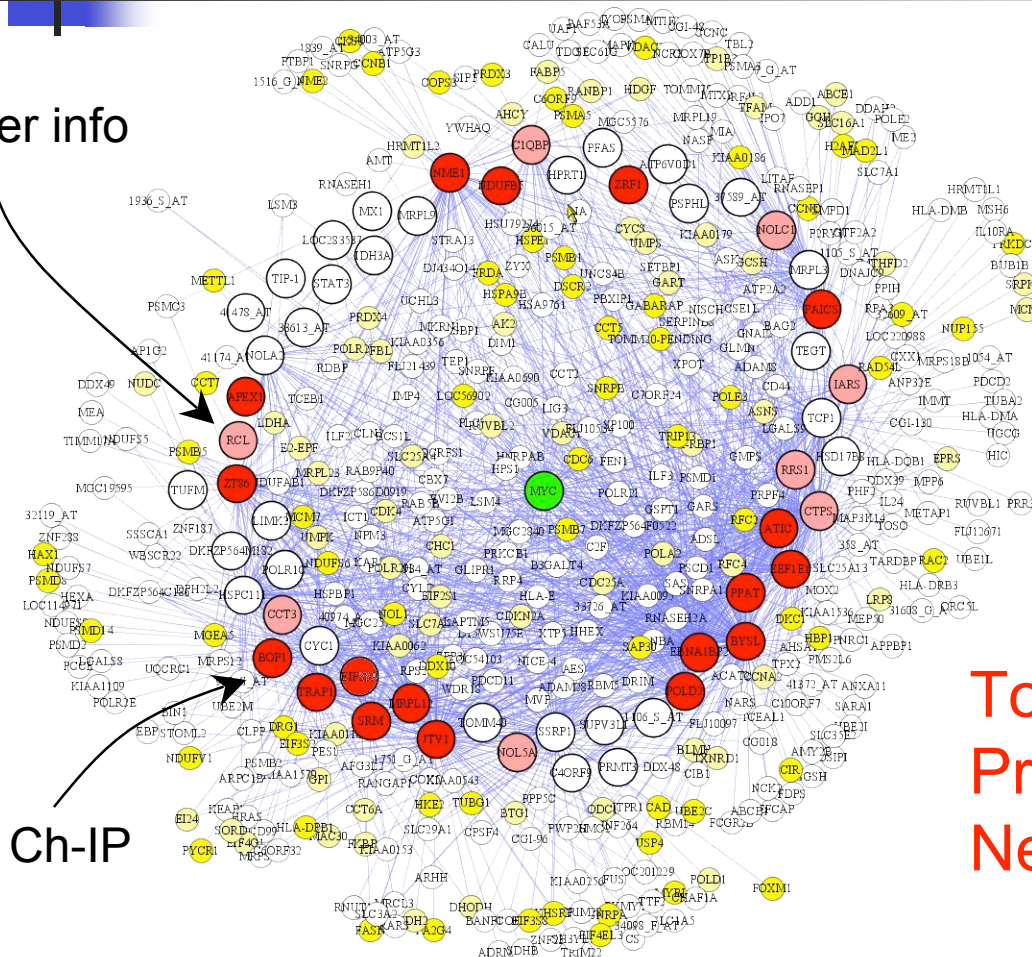
# B-cell dataset

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- ~400 arrays
- No dynamics
- ~250 naturally occurring, ~150 perturbed
- ~25 phenotypes (normal, tumors, experimental perturbations)
- Expression range due to differential expression in different phenotypes

# c-MYC subnetwork

other info



Ch-IP

- Protooncogene,
- 12% background binding,
- one of top 5% hubs
- significant MI with 2000 genes

Total interactions: 56  
Pre-known: 22  
New Ch-IP validated: 11/12



# Problem:

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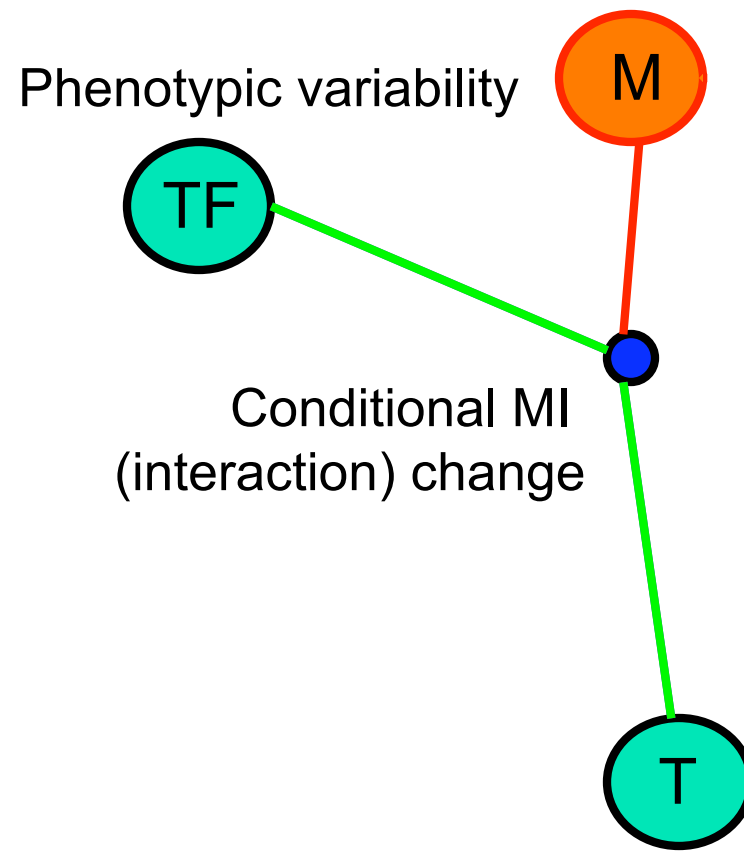
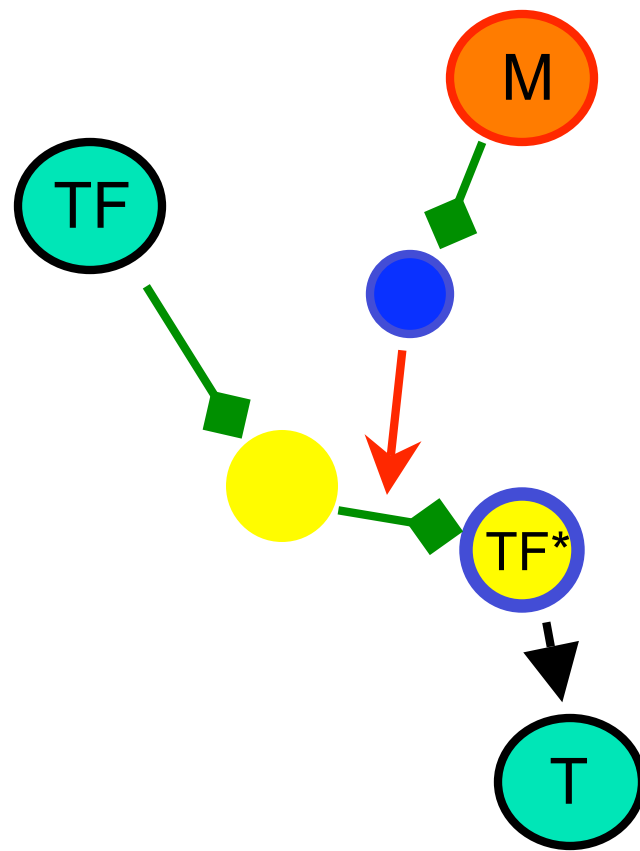
- Much of regulation in higher eukaryotes is post-transcriptional (e.g., splicing), and post-translational (e.g., phosphorylation, complex formation).
- Many mRNA (e.g., p53) constitutively expressed.

Can these be observed from mRNA expressions only?

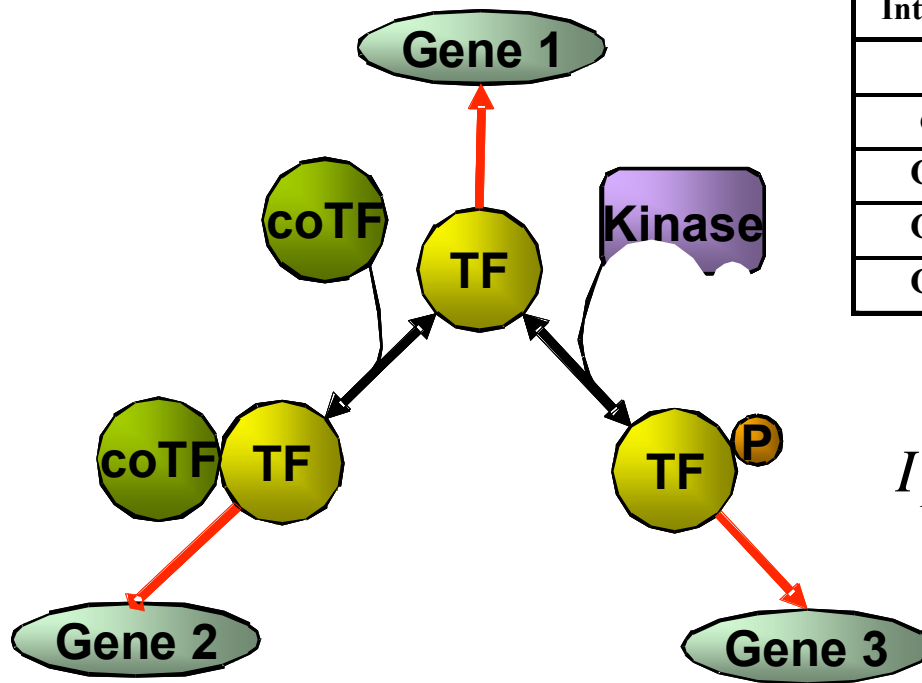
## Solution:

Phenotypic and population variability (even in constitutively expressed genes) induces higher order dependencies between TFs, targets, and modulators.

# Nontranscriptional modulation from mRNA expression



# Numerical case study: Transistor modulation

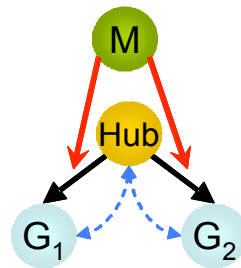


TF Interaction	$I$	$I_{PK}^-$	$I_{PK}^+$	$\Delta I_{PK}$	$I_{coTF}^-$	$I_{coTF}^+$	$\Delta I_{coTF}$
PK							
coTF							
Gene1	0.73	0.54	0.57		0.55	0.54	
Gene2						0.37	0.37
Gene3			0.35	0.34			

$$I_{PK}^+(\text{gene1}) = I(TF, \text{gene1} \mid PK \text{ high})$$



# Enforcing irreducibility: ARACNE on a TF-hub

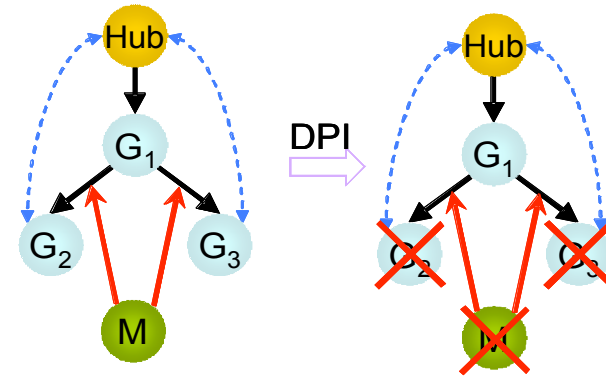
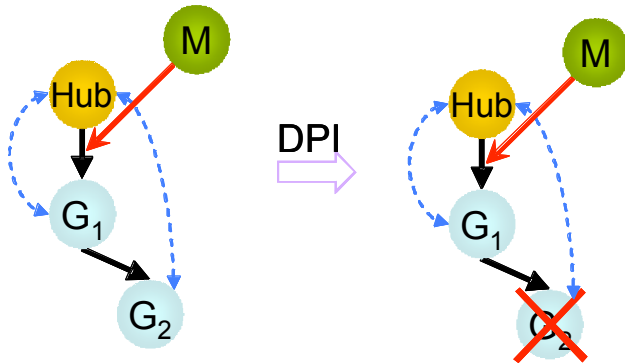


LEGEND:

→ Transcriptional regulation

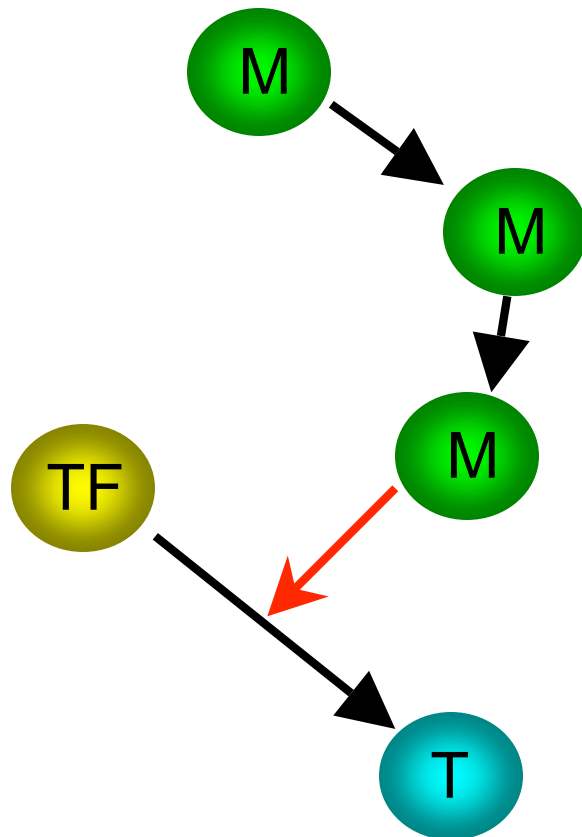
→ Post-transcriptional modulation

↔ Significant  $\Delta I$  conditioned on M



However:

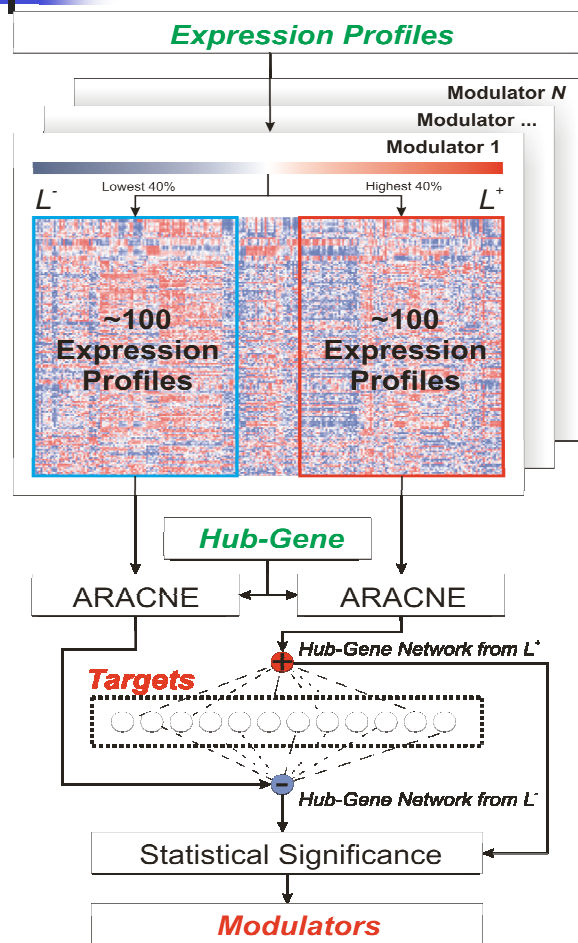
No solution yet for...



Modulators are not  
irreducible.

Any suggestions?

# Algorithm flowchart



- Focus on a hub (c-MYC).
- Select modulators with  $\sigma > \text{microarray noise}$  (Tu et al., 2002) -- many signaling genes, constitutively expressed genes.
- Find modulators whose expression inflicts **significant conditional MI changes** for an ARACNE target in at least one conditional topology.
- **No guarantee of modulator irreducibility.**
- **Guarantee of target irreducibility** (after multiple hypothesis correction).

$$\Delta I(g_{TF}, g_t | g_m) =$$

$$= \left| I(g_{TF}, g_t | g_m^+) - I(g_{TF}, g_t | g_m^-) \right| > 0$$

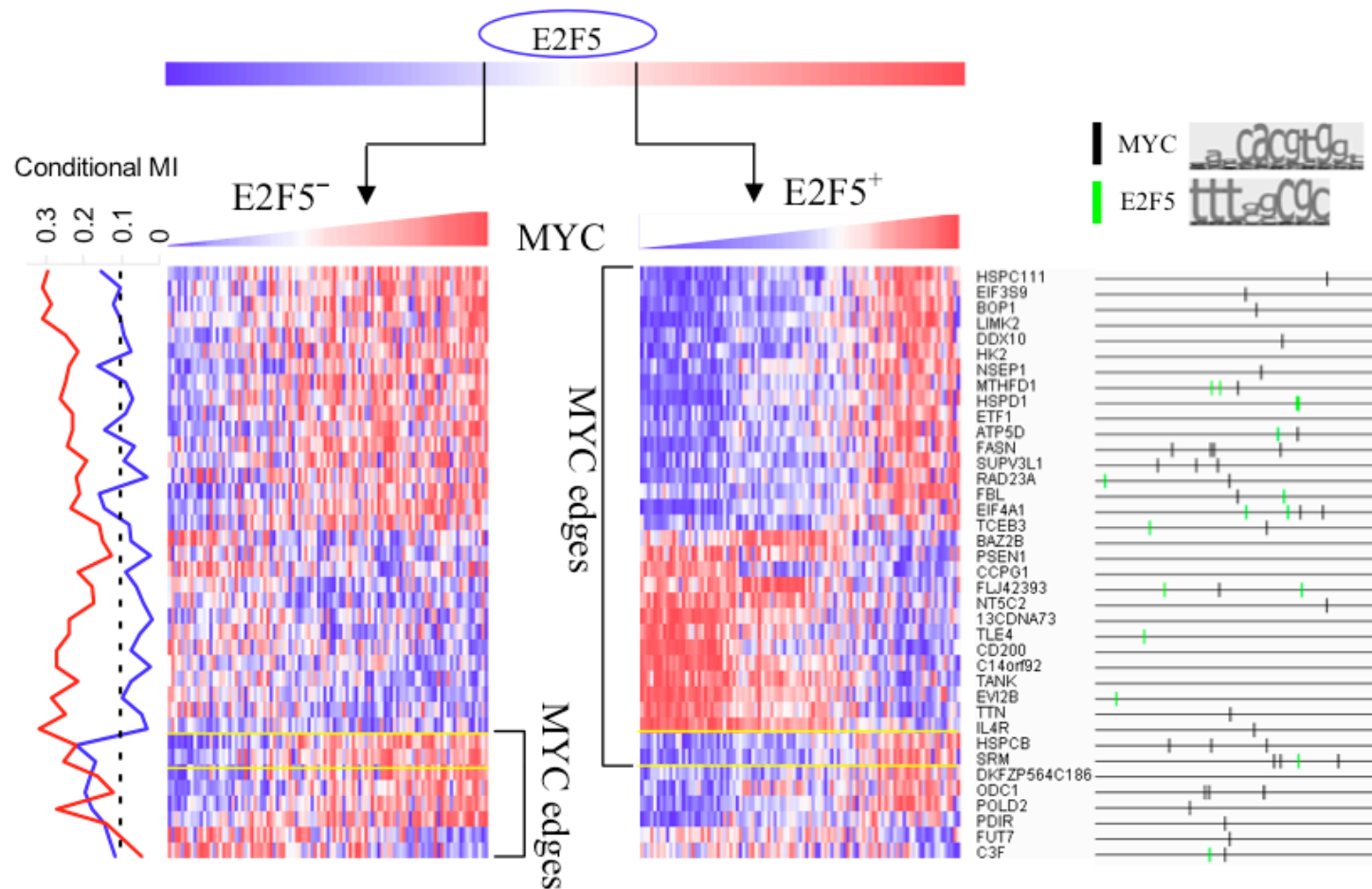


# c-MYC modulators

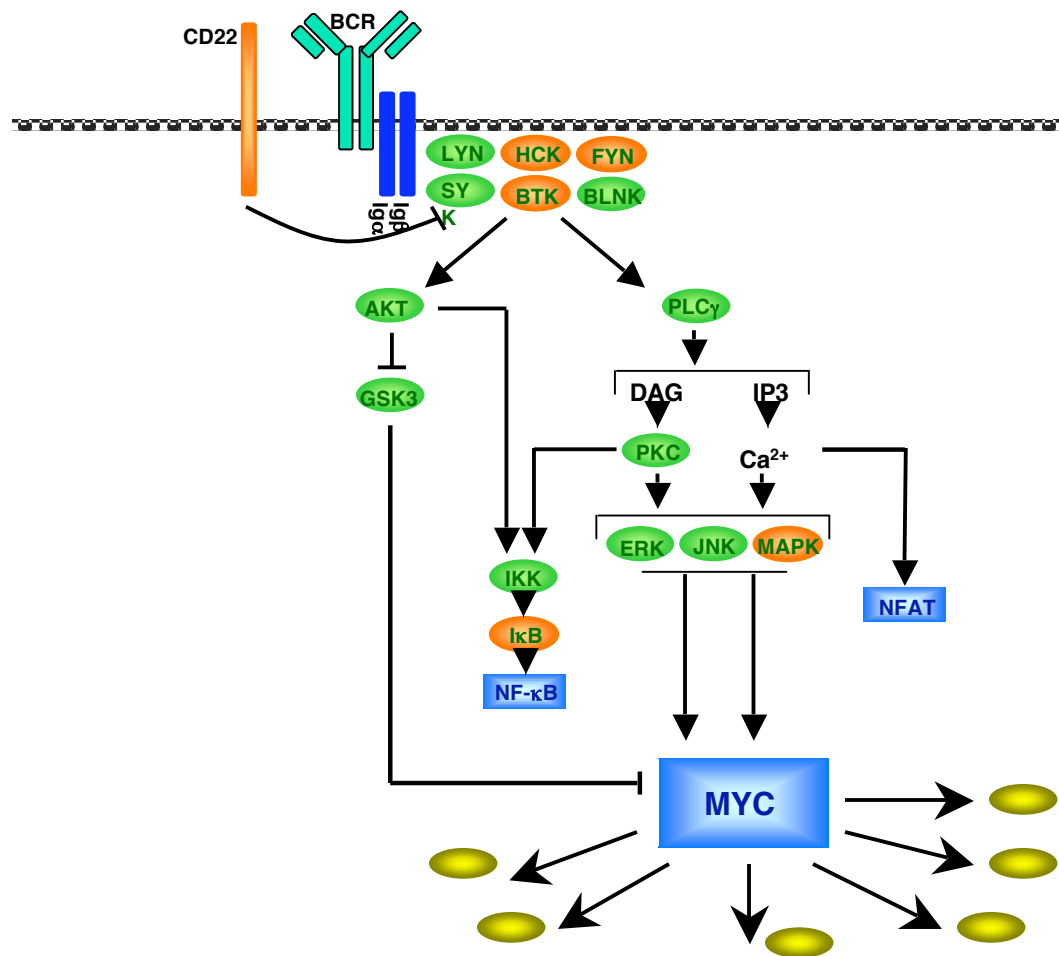
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- 1117 candidate modulators
- 100 modulators, 130 targets, 205 interactions
- GO enrichment of the modulator set: kinases, acyltransferases, TFs (all  $p < 5\%$ )
- Modulators in known MYC regulation pathways (e.g., BCR)
- TFs: 15/100,  $p = 1e-6$ .
- 4/5 TF modulators (e.g., E2F5) with TRANSFAC signatures have binding sites in modulated targets promoter regions.
- Modulators with many ( $\geq 4$ ) targets are not-specific (proteolysis, upstream signaling components, receptor signaling molecules).
- Modulators with few (1-2) effected targets are mostly co-TFs, interaction-specific.
- $\sim 1/3$  modulators are literature-validated.
- Biochemical validation of predictions in progress.

# Example: TF co-factor modulator



# BCR pathway: Reducibility



- predicted modulators
- not in the candidate list
- TF's not predicted
- Protein complex
- Targets